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| **A close up of a logo  Description automatically generated**  **ANIMAL CARE AND USE PROTOCOL**  **For Teaching**  **Mahidol University-Institute Animal Care and Use Committee**  **(MU-IACUC)** |

**COVER SHEET**

**Overview**

This section will be completed by the MU-IACUC

|  |  |
| --- | --- |
| **Protocol number** | **F05 -** |
| **Date of submission** (dd/mm/yy) |  |
| **Date of Request modification** (dd/mm/yy) |  |
| **Date of Resubmitted** (dd/mm/yy) |  |
| **Date of Approved/Disapproved** (dd/mm/yy) |  |
| **Date of Expiration** (dd/mm/yy) |  |

**1. Subject:**

(Thai)

(English)

**Course** **Course Title**

Study level

(\*Please attach course syllabus)

**2. Course coordinator:** *The names of each person, under this section, will be appeared on the “Certificate of Approval”.*

Name

Position: Department Faculty/Institute

Tel. E-mail

\* Animal use license no Expired date

*\*Issued by Institute of Animal for Scientific Purposes Development, NRCT*

**3.** **Contact person in case of emergency**:

Name

Position: Department Faculty/Institute

Work phone Mobile phone

E-mail

**6.** **Duration of Protocol:**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Period for which the protocol is required  (must not exceed three years) | | Years |  | Months |  |
| Start date |  | End date | |  | |
| Please submit your application one to two months (preferably two months) before your planned start date. | | | | | |

(The start date has to be after the date of application submission. Please note that no animal use may occur until the Animal Ethics Committee approves, and all animal use must be finished before the end date. The date format is dd/mm/yyyy.)

*\*Animal care and use protocol for teaching certification period is up to 3 years depend on decision of MU-IACUC]*

**5. Signatures** *Your signature as P.I., Co-investigator on this application verifies that the information herein is true and correct and that you are familiar with and will comply with standard of animal care and use established under the ethical guidelines and policies of the Mahidol University and Office of the National Research Council of Thailand (NRCT) and the Animal for Scientific Purpose Act., B.E. 2558*

**Course coordinator:** Date  **( )**

***The signature of Dean of Faculty / Head of Institute verifies that he / she acknowledges the fact that P.I. under this affiliation will conduct the animal care and use protocol, as provided herein.***

**Head of Faculty/Institute:** Date **( )**

**Faculty/Institute**

#### MAHIDOL UNIVERSITTY

#### RESEARCH PROTOCOL FORMAT

#### FOR PERMISSION OF ANIMAL CARE AND USE

**Type of animal protocol**

Full Protocol  Exemption Protocol (answer only item 1-4, 6-8, 9.1.1and 22-24)

**1. Course description**: *(Provide a brief, only one A4 page, and simplified description of the project that is easily understood by non-scientists, expressing its significance and needs for undertaking the study).*

**2. Rationale and literature review:** *(Include a brief statement of the requirement for the information being sought. Typically, the literature or the experience that led to the proposal will be briefly reviewed, references cited will be provided)*.

**3. Reference search:***(Please specify the references used for researching the course details)****.***

**4. Objective(s):** *(Provide goal/specific aim of this project)*

**5. Potential benefits of the study:** *Explain how the study is important to human or animal health and the advancement of knowledge*

**6. Teaching design and animal procedures:** *Provide a complete, step-by-step description of the experiment(s). Describe in detail the experimental procedures especially what will be done from obtaining the animals to the end of animal experiment(s). Diagram(s) or flow chart(s) should accompany complex experimental design*.

**7. Data analysis and statistical methods:** *Describe statistical methods to be used for analysis of the results and for testing the hypothesis*

**8. Animal care and Source of samples or specimens:**

**8.1 Provide description of animals in Table below**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Common name | species | Strain/ Stock | Age | Weight | Sex | Number |
|  |  |  |  |  |  |  |
|  |  |  |  |  |  |  |
|  |  |  |  |  |  |  |

**8.2 Students / animal**

8**.3 Permanent animal ID method:** *(eg. ear tag, ear punch, microchip, tattoo, N/A, other please specify)*

**8.4 Special consideration:** *(List specialized requirements for the research animals, if any)*

**8.5 Source/Vendor:**

|  |  |  |  |
| --- | --- | --- | --- |
| From previously approved protocol | | | Protocol number (*please specify*): |
| Protocol Title (Thai) *please specify*: | | | |
| Protocol Title (English) *please specify*: | | | |
| Nature  (*If From wildlife must be complied with the Wildlife Preservation and Protection Act B.E.2562(2019) and National Parks Act B.E.2562(2019)*, **Please attach the permission document)** please specify: | | | |
| **Laboratory animals supply**  (With genetic quality and health certificates) | | please specify: | |
| Other | please specify: | | |

**8.6 Explain why the proposed animal species is/are the most appropriate**

**8.7 Provide a statistical analysis for estimation of sample size with an explanation for the number of animals to be used**

8**.8** **Transportation** (if any,please specify how will the animals be transferred to the lab)

8.**9 Prevention of injury and/or infection**

8**.10** **Is the quarantine required?**

No

Yes, *specify the method, location and duration*

**9. Animal care:**

**9.1 Husbandry consideration**: *(Briefly describe animal housing and living conditions, routine animal observations, feed and water provisions, etc.)*

9.1.1 Study location (specify room number, name of building or facility)

9.1.2 Housing system:

Clean conventional  Strict hygienic conventional

Isolator maintained  Barrier maintained

Laminar flow

Other, please specify

9.1.3 Caging:

Solid bottom, open top  Static filtered top cages

Suspended cages, wire bottom  Metabolic cages

Individual ventilated cage (IVC)

Other, please specify

9.1.4 Cage size: W x L x H, (inch)

9.1.5 Caging materials:

Plastic  Stainless steel

Other, please specify

9.1.6 Number of animals per cage:

9.1.7 Social housing (more than one animal per cage):   
(*The IACUC requires social housing of all social animals)*

Yes  No

If NO, provide scientific justification for not socially housing the animals. Describe what will be done to replace this social contact with conspecifics.

9.1.8 Environmental requirements:

Temperature:

Humidity:  30 – 70 % relative humidity,  Other, specify

Light:  Standard fluorescent,  Other, specify

Light cycle:  Standard (12:12 hours),  Other, specify

9.1.9 Food

Type of food:  Standard diet,  Other, specify

Feeding schedule:  Routine feeding (ad libitum),  Other, specify

9.1.10 Water

Type of water:  Reverse osmosis,  Other, specify

Provision of water:  Routine feeding (ad libitum),  Other, specify

9.1.11 Bedding

No

Yes, please specify  Sterile  Non-sterile

Type of bedding:  Wood shaving  Sawdust

Paper  Other, specify

Schedule of changing:  Once a week,  Other, specify

9.1.12 Environmental Enrichment:

Accept

Decline, provide scientific justification

9**.2 Is this project intended to conduct the animal experiment in other building?**

*(This is allowed for conducting experiment(s) only not for housing. In addition, the holding period must be less than 12 hours).*

No  Yes, *answer all that apply in* ***9.2.1*** *to* ***9.2.4***

**9.2.1** Where the experiment is expected to be conducted? Please indicate the building name and room number.

**9.2.2** Please provide the animal experimental procedures in detail.

**9.2.**3 Estimated total time period that live animals will be kept in the laboratory is hours

**9.2.**4 How will the animal sample or carcass be disposed?

**10. Veterinary medical care**: *(Describe the routine veterinary care. List the criteria used for health evaluation while the animals are on study).*

**11. Animal welfare:**

**11.1 Replacement, Reduction and Refinement**. (*Briefly describe how you have considered each of the following alternatives (the 3Rs) or why they are not applicable).*

11.1.1 Replacement of animals *(e.g., with in vitro models, computer models or less sentient animals)*

11.1.2 Reduction in the number of animals *(e.g., using appropriate statistical methods in the design and analysis of the study; reduction in experimental variability by using animals of defined genetic or microbiological status)*

11.1.3 Refinement of experimental procedures to minimize pain or distress*(e.g., early endpoints; use of analgesics, anesthetics or sedatives; techniques that reduce stress in the animal.)*

**11.2 Potential animal pain and distress assessment:**

11.2.1Please indicate pain category according to USDA Pain and Distress.(Appendix A)

**Category B:** *Animals being bred or housed without any research manipulations or non-invasive observation of animals in the natural habitat*

Number of animals

**Category C:** *Animal use activities that involve no more than momentary or slight pain or distress (no greater than an injection) where there is no need for use of pain-relieving drugs*

Number of animals

**Category D:** *Animal use activities that involve accompanying pain or distress to the animals and for which appropriate anesthetics, analgesics, tranquilizing drugs, and/or humane endpoints are used to avoid pain, distress, or discomfort*

Number of animals

**Category E:** *Animal use activities that involve accompanying pain or distress to the animals and for which appropriate anesthetic, analgesic, tranquilizing drugs; or other methods for relieving pain or distress are NOT used*

Number of animals

*Provide strong scientific justification as to why pain-relieving drugs or other methods for relieving pain cannot be used on animals.*

11.2.2 During the study:

1) How often will the clinical condition of animals be monitored?

2) Who will monitor the clinical condition of the animals?

11.2.3 Are the animals expected to experience any **specific study-induced or related problems** (i.e. health problems, pain, distress, complications, etc.) **or any health problems** as a result of the phenotype of the animal?

No  Yes, *answer all that apply in 11.2.3.1 to 11.2.3.2*

11.2.3.1 Describe the expected problems.

11.2.3.2 What criteria(s) will be used to assess pain, distress, or discomfort? Check all that apply:

Inactivity

Loss of appetite

Loss of weight  5%  10 %  15%  20% weight loss

Restlessness

Abnormal resting postures, somnolence or hunched posture

Licking, biting, scratching, or shaking a particular area

Failure to show normal patterns of inquisitiveness

Failure to groom, causing and unkempt appearance

Guarding (protecting the painful area)

Loss of mobility

Red stain around the eyes of rats

Self-mutilation

Labored breathing

Tumor

Unresponsiveness

Other (please list)

11.2.4 Literature search for alternative to procedure that cause pain & distress

11.2.4.1 Database(s) searched*(Please specify the database name, e.g., PubMed, ScienceDirect))*:

11.2.4.2 Date of literature search (must be within six months prior to submission date) *(dd/m/yy))*

11.2.4.3 Range of years searched *(To prevent the duplication of your proposed experiment, the minimum period of search should be more than 5 years)*

11.2.4.4 Key words used in search:

11.2.4.5 Results of search: Does the proposed research duplicate any previous work?

No  Yes

If YES, explain why it is scientifically necessary to duplicate previous experiment.

**11.3 Anesthesia**

No  Yes, please answer the following questions:

1) Preanesthetic preparation:

2) Anesthetic agent(s) used:

3) Dosage:

4) Volume:

5) Route of administration:

6) Frequency of anesthesia:

7) Length of anesthesia:

8) Who is responsible for monitoring anesthesia?

9) If an inhalation anesthetic is used, describe scavenging of the waste anesthetic gas.

10) What criteria(s) will be used to assess level of anesthesia?

Check all that apply:

Respiration rate  Body temperature  Heart rate

ECG  Toe pinch  Tail pinch

Corneal reflex  Pedal reflex  Muscular relaxation

Color of mucous membrane

Other (pulse oximeter, respirometer) please list

11) How animals are kept warm?

**11.4 Analgesics and/or tranquilizers:**

No  Yes, please specify

1. 1) Type of analgesics used
   * + 1. Agent(s)
2. 2) Dosage

3) Route of administration

4) Schedule

**11.5 Describe post-anesthetic treatment or intervention:**

**12. Surgery:**

No —*proceed to 13*  Yes, *answer all that apply in 12.1 to 12.7*

**12.1 Surgical procedure** is:  Non-survival  Survival

Major  Minor

One time  Multiple

**12.2** **Location**: *(Give the location/room number for the proposed surgical procedure.)*

**12.3** **Surgeon/qualification***: (Indicate who will perform the surgery, and his/her qualifications, training, or experience in the proposed procedure*.*)*

**12.4 Procedure**: *(Describe in detail the surgical procedure*.*)*

**12.5** **Pre- and post-operative provision**: *(Detail the provision for both pre-and*

*post-operative care, including provisions for post-surgical observation.)*

**12.6 Describe long-term care of chronic survival procedure**.

**12.7** **Multiple survival surgery procedures**: *(Multiple major operative procedures on the same animal must be adequately justified for scientific reasons by the principal investigator in writing.)*

12.7.1 Procedure:

12.7.2 Scientific justification:

12.7.3 Who will be the responsible for post-surgical care and treatment?

**13. Blood or body fluid withdrawal/tissue collection/injections, tail clip, Gavage**

*(Describe in detail: method(s), needle size(s), volume(s) collected or administered, and frequency of collection or injection.)*

| **Procedures** | **Anatomic**  **location** | **Needle size/**  **catheter size**  **and length** | **Biopsy**  **size** | **Volume**  **collected**  **(ml)** | **Volume**  **administered**  **(ml)** | **Frequency** |
| --- | --- | --- | --- | --- | --- | --- |
| **Blood withdrawal** |  |  |  |  |  |  |
| **Body Fluid**  **withdrawal** |  |  |  |  |  |  |
| **Tissue collection** |  |  |  |  |  |  |
| **Injection** |  |  |  |  |  |  |
| **Infusion** |  |  |  |  |  |  |
| **Tail clip** |  |  |  |  |  |  |
| **Gavage** |  |  |  |  |  |  |
| **Other *(specify)*** |  |  |  |  |  |  |

**Total blood volume ml. in total study days or**  **months**

**14. Use of non-pharmaceutical grade compounds**

**14.1 Will animals be treated with non-pharmaceutical grade compounds?**

No —*proceed to 15*

Yes, *answer all that apply in 14.2 and 14.3*

**14.2 Give information on name, source, formulation, concentration, site and route of administration and potential side effects**

**14.3 Provide scientific justification for the use of non-pharmaceutical grade compounds**

**15. Restraint with mechanical devices:**

No—*proceed to 16*   Yes, *answer all that apply in 15.1 and 15.2*

**15.1 Describe device, duration of restraint, frequency of observation, conditioning procedures and steps to assure comfort and well-being.**

**15.2 Provide scientific justification for prolonged complete restraint**

**16. Project involving food and water deprivation, or dietary manipulation:**

**16.1 Does this protocol involve food or water deprivation or dietary manipulation?**

No —*proceed to 17*

Yes, *describe methods for assessing physical conditions (e.g., weight loss), pain, discomfort and stress during the course of study. Include clinical signs and symptoms expected.*

**16.2 Provide detail of these procedures in Table below**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Procedures** | **Amount restricted or added** | **Duration** | **Compound supplemented** | **Compound excluded** | **Frequency** |
| **Food deprivation** |  |  |  |  |  |
| **Fluid deprivation** |  |  |  |  |  |
| **Nutrient alteration** |  |  |  |  |  |

**17. Tumor study, use of disease models and toxicity testing**

**17.1 Does this protocol involve tumor study, use of disease models or toxicity testing?**

No—*proceed to 18*

Yes, *answer all that apply in 17.2 and 17.3*

**17.2 Describe methods for assessing physical conditions, stress, pain and discomfort during the course of study. Include clinical signs and symptoms expected.**

**17.3 What are the criteria for humane endpoint in this protocol?**

**18. Behavioral studies:**

**18.1 Does this protocol involve behavioral study?**

No—*proceed to 19*

Yes, *answer all that apply in 18.2 to 18.3*

**18.2 Describe type of behavioral manipulation**

**18.3 Describe the protocol involving the use of testing apparatus or aversive stimulus and detail of duration and frequency of the testing period**

**19. Study endpoints**

**19.1 Describe the endpoint for the animals in this protocol**. *Indicate whether recovery, euthanasia, or death is/are expected, and when the animal experimentation phase will be stopped.*

**19.2 Humane (early) endpoint is used** *(i.e., animals are humanely euthanized prior to the expected day of termination)*

No

Yes, *provide criteria for humane endpoint*

**19.3 Death or moribund as an endpoint is used**

No

Yes, *answer all that apply in 19.3.1 to 19.3.2*

19.3.1 Provide criteria that establish when this endpoint has been reached, and describe how animals will be monitored and cared for

19.3.2 List persons responsible for evaluating animal condition, record keeping, and notifying PI and/or veterinarians to perform euthanasia

**20.** **Animal euthanasia and disposition**

**20.1 After completion of activity, the animals will be:**

Euthanized

Returned to production/breeding unit/facility inventory

Transferred to another research project:

Protocol No. and name of principal

Other, specify

**20.2** Describe **the two-step euthanasia method** according to AVMA guidelines

This protocol uses:

Step 1 - Describe chemical method:

Immersion:

Pharmaceutical-grade clove oil at ml/L for minutes

MS-222 at mg/L for minutes

Magnesium salts at mg/L for minutes

Ethanol at % ml/L for minutes

2-phenoxyethanol at ml/L for minutes

Benzocaine at mg/L for minutes

Lidocaine at mg/L for minutes

Isoflurane at mg/L for minutes

Sevoflurane at mg/L for minutes

Quinaldine sulfate at mg/L for minutes

Other, specify

Injection:

Pentobarbital at mg/kg

Ketamine at mg/kg

Ketamine-medetomidine at mg/kg

Other, specify at mg/kg

Route of administration

Intramuscular,  Intravenous,  Intraperitoneal,  Intracoelomic,   
 Intracardiac

Step 2 - Describe mechanical method:

Cervical transection

Chilling using:  Ice,  Cold water for minutes

Cranial concussion

Decapitation

Exsanguination

Maceration

Penetrating captive bolt

Pithing

Other mechanical method, specify

Use another method besides mechanical method, *describe and provide strong scientific justification*

**20.3 State how death will be verified before disposal:**



**21. Necropsy/ Selected tissue and sample collection**

No

Yes, *provide room number, personnel with qualification*

**22. Animal sample utilization and disposal:**

**22.1 Subsequent use of animal samples,** when the primary research is completed, are any remaining animal samples used for other proposes?

No

Yes, *please describe the specific parts and purpose(s), such as archival for future studies, inclusion in a teaching collection, donation to a museum, etc.*

**22.2 Animal tissue and carcasses disposal:** *Describe method used to dispose animal tissue and carcasses.*

**23.** **Occupational health and safety:**

**23.1 Select types of hazards associated with this protocol,** also provide name, source and amount to be used in each category

Cancer cell lines

Infectious agents*provide the certificate of biosafety approval*

Hazardous chemicals (e.g., carcinogen, mutagen and teratogen)

Radiation equipment and radioactive elements

Recombination agents

Other, specify

None

**23.2 Specify biosafety level:**  BSL-1  BSL-2  BSL-3

**23.3 Explain how the wastes associated with these hazards are decontaminated and disposed**

**23.4 Explain how the carcasses associated with these hazards are disposed**

**23.5 Explain any safety precautions and protective measures** (e.g., biosafety cabinet and proper PPE) to protect personnel from those hazards and list any surveillance procedures in place to monitor any potential exposure

**23.6 In case of accident, provide immediate procedures and early treatment to limit possible injury or illness**

**24. Qualification of teaching staff:**

*List all individuals who will be* ***directly*** *involved with the animals on this project. Include all investigators, students, post-doctoral researchers, staff research associates and laboratory assistants who will actually work with the animals. If personnel do not have experience, state how they will be trained:*

|  |  |  |  |
| --- | --- | --- | --- |
| **Name** | **Responsibilities** | **Direct involvement with animal samples (%)** | **Relevant experience and qualification**  **(e.g. How many years of experience working with animals or training related to the research)** |
|  |  |  |  |
|  |  |  |  |

*As Principal investigator on this protocol, I verifies that the information herein is true and correct and that I am familiar with and will comply with standard of animal care and use established under the ethical guidelines and policies of Mahidol University, and Office of the National Research Council of Thailand (NRCT). Additionally, I acknowledge my responsibilities and provide assurances for the followings:*

**A. Animal use:** The animals authorized for use in this protocol will be used only in the activities and in the manner described herein, unless a deviation is specifically approved by the MU-IACUC.

**B. Duplication of effort:** I have made a reasonable, good faith effort to ensure that this protocol is not an unnecessary duplication of previous experiments.

**C. Statistical assurance:** I assure that I have consulted with qualified statistician to evaluate the statistical design or strategy of this proposal, and that the minimum number of animals needed for scientific validity are used.

**D. Biohazard/safety:** I have taken into consideration, and I have made the proper coordination’s regarding all applicable rules and regulations concerning radiation protection, biosafety, recombinant issues, etc., in the preparation of this protocol.

**E. Training:** I verify that the personnel performing the animal procedures/manipulations described in this protocol are technically competent and have been properly trained to ensure that no unnecessary pain or distress will be caused as a result of the procedures/manipulations.

**F. Responsibility:** I acknowledge the inherent moral and administrative obligations associated with the performance of this animal use protocol, and I assure that all individuals associated with this project will demonstrate a concern for the health, comfort, welfare, and well-being of the research animals. Additionally, I pledge to conduct this study in the responsibility for implementing animal use alternatives where feasible, and conducting humane and lawful research.

**G.** **Scientific review:** This proposed animal use protocol has received appropriate peer scientific review, and is consistent with good scientific research practice.

Course coordinator Date

( )